Intermittent hypoxia training protects cerebrovascular function in Alzheimer's disease

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Abstract

Alzheimer's disease (AD) is a leading cause of death and disability among older adults. Modifiable vascular risk factors for AD (VRF) include obesity, hypertension, type 2 diabetes mellitus, sleep apnea, and metabolic syndrome. Here, interactions between cerebrovascular function and development of AD are reviewed, as are interventions to improve cerebral blood flow and reduce VRF. Atherosclerosis and small vessel cerebral disease impair metabolic regulation of cerebral blood flow and, along with microvascular rarefaction and altered trans-capillary exchange, create conditions favoring AD development. Although currently there are no definitive therapies for treatment or prevention of AD, reduction of VRFs lowers the risk for cognitive decline. There is increasing evidence that brief repeated exposures to moderate hypoxia, i.e. intermittent hypoxic training (IHT), improve cerebral vascular function and reduce VRFs including systemic hypertension, cardiac arrhythmias, and mental stress. In experimental AD, IHT nearly prevented endothelial dysfunction of both cerebral and extra-cerebral blood vessels, rarefaction of the brain vascular network, and the loss of neurons in the brain cortex. Associated with these vasoprotective effects, IHT improved memory and lessened AD pathology. IHT increases endothelial production of nitric oxide (NO), thereby increasing regional cerebral blood flow and augmenting the vaso- and neuroprotective effects of endothelial NO. On the other hand, in AD excessive production of NO in microglia, astrocytes, and cortical neurons generates neurotoxic peroxynitrite. IHT enhances storage of excessive NO in the form of S-nitrosothiols and dinitrosyl iron complexes. Oxidative stress plays a pivotal role in the pathogenesis of AD, and IHT reduces oxidative stress in a number of experimental pathologies. Beneficial effects of IHT in experimental neuropathologies other than AD, including dyscirculatory encephalopathy, ischemic stroke injury, audiogenic epilepsy, spinal cord injury, and alcohol withdrawal stress have also been reported. Further research on the potential benefits of IHT in AD and other brain pathologies is warranted.

Keywords: Alzheimer's disease, brain ischemia, cerebral circulation, cerebrovascular risk factors, cognitive function, intermittent hypoxia training, neurodegeneration, nitric oxide, nitrosative stress, oxidative stress, vasoprotection

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Introduction

Alzheimer's disease (AD) is characterized by deteriorating executive function, verbal fluency, psychomotor speed, and mental flexibility, which progress along with neurodegeneration to produce severe cognitive impairment, dementia and, eventually, death.^{1,2} The histological hallmarks of AD include the extracellular accumulation of two amyloid- β (A β) peptides, A β 1-42 in the form of neuritic "senile" plaques in close association with astroglia and A β 1-40 deposits in the perivascular spaces around cerebral arteries and microvessels, and neurofibrillary tangles of hyper-phosphorylated tau within the cell bodies of neurons.³⁻⁶ The most severe AD pathology is found in the hippocampus, amygdala, and mediotemporal regions, followed by the

temporo-parieto-occipital junction. AD also affects the frontal heteromodal cortex, temporal cortex, and the basal nuclei.^{7,8} AD is incurable; current interventions at best slow its progression, but can neither arrest nor reverse the neurocognitive impairment.²

AD is a looming epidemic, especially in developed societies with extended longevity. In the United States alone, approximately 5.2 million people had dementia in 2010, and that number is expected to increase to 13.2 million by 2050.⁸ Worldwide, 36 million people had dementia in 2010; by 2050, it is predicted that a staggering 115–140 million people will have dementia.^{9–11} Of these dementia patients, 60–80% have AD, and most of the remainder have a closely related syndrome, vascular dementia. In addition to the toll on the patients and their loved ones, AD has become the third costliest disease entity in the United States, exceeded only by cardiovascular disease and depression.¹¹

A small minority of AD cases is categorized as hereditary or familial AD.^{2,12} Familial AD can strike earlier in adulthood, when patients are in their 40s and 50s. Familial AD carries a grim prognosis, with rapid onset and progression of the dementia culminating in death within 2–3 years of the onset of overt symptoms. Idiopathic (sporadic) AD typically afflicts patients later in life, and its incidence mounts beyond age 65. An estimated 40% of Americans aged \geq 85 years have clinically significant cognitive impairment, of which 75–80% have AD.¹³ AD is the fifth-leading cause of death among Americans \geq 65 years old,¹⁴ and the incidence of AD doubles every five years beyond age 65.⁹

Sporadic AD is associated with a variety of risk factors (Figure 1). Unmodifiable factors include advanced age, expression of the apolipoprotein E4ɛ (APO-E4ɛ) isoform, and traumatic brain injury.¹³ The APO-E4ε gene has been identified as a major risk factor for AD; indeed, 40-50% of AD patients are homo- or heterozygous carriers of the APO-E4ɛ allele.¹³ In addition, several modifiable AD risk factors have been identified, including cigarette smoking, hypertension, type 2 diabetes mellitus, physical inactivity, dyslipidemia, adiposity, obstructive sleep apnea, and metabolic syndrome.^{15,16} Since each of these modifiable risk factors has a cardiovascular component,¹⁷ collectively they comprise vascular risk factors for AD, i.e. VRF.^{1,18} It is not surprising that cerebral underperfusion in AD has been reported,^{19,20} although the cause-and-effect relationship between cerebrovascular dysfunction and AD is a matter of intense debate.²¹

This review will focus on interactions between cerebrovascular function and development of AD. In particular, we will discuss factors responsible for the depression of cerebral blood flow and its contributions to AD. In this regard, we will review evidence that reducing VRF delays AD or slows its progression. In addition, evidence will be presented that moderate, intermittent hypoxia training (IHT) reduces VRF. Finally, we will examine recent preclinical evidence that IHT directly improves cerebral perfusion and cognitive function in animals with experimental AD. Although there are no reports of IHT treatment of humans with AD, reports showing beneficial effects of IHT in other human brain pathologies will be discussed. This review includes Eastern European studies of IHT that are not generally well known in the West.

Cerebrovascular function and AD

The human brain's intense energy requirements demand continuous delivery of O_2 and metabolic substrates to support ATP production. Like the left ventricular myocardium,²² the brain has a relatively high perfusion rate and limited oxygen extraction reserve that vary temporally and spatially according to specific sensory inputs and regional activity.²³ To meet changing regional requirements for oxygen and metabolic fuels, the cerebral circulation must adjust its conductance and does so by metabolic autoregulation.²⁴ In the normal cerebral circulation, regional changes in brain function and associated demand for oxygen are readily met by local arteriolar dilation resulting from cellular release of vasodilatory metabolites, including nitric oxide (NO), carbon monoxide, adenosine, and opioidergic



Figure 1 Alzheimer's Disease (AD) risk factors. Risk factors for AD are grouped into those that cannot be modified, i.e. are fixed (upper left) and those that are modifiable by intermittent hypoxia training (IHT) and potentially other interventions (upper right). These fixed and modifiable risk factors give rise to the injury mediators amyloid β , inflammation, reactive oxygen and nitrogen species (RONS), and suppression of endothelial nitric oxide synthase (eNOS) which, in turn, damage macro- and microvascular components of the cerebral circulation. The resultant impairment of cerebral blood supply causes neurodegeneration which culminates in neurocognitive impairment

peptides²⁵⁻²⁷ associated with altered tissue PO₂.²⁸ However, atherosclerotic disease in the cerebral supply arteries limits the vascular reserve that is required for autoregulation of cerebral blood flow and exposes the otherwise healthy, active brain to the potential damage of sustained hypoxia and ischemia.^{29–31} The cerebral circulation must also effectively remove metabolic wastes, as well as toxins produced by cerebral disease, most notably $A\beta$, which accumulates in the brain's perivascular interstitial spaces when cerebrovascular function is compromised.³² Studies of radiolabeled proteins in rats suggest that pulsations of the cerebral microvasculature secondary to pulsatile flow effect Aß removal by propelling the peptide along the perivascular interstitium into the lymphatic drainage.³³ Accordingly, Weller et al.³³ proposed that vascular stiffening, by dampening pulsatile flow, impairs perivascular AB clearance, culminating in cerebral amyloid angiopathy.

In addition to atherosclerotic disease, disorders of cerebral small arteries, arterioles, and capillaries are increasingly recognized as a threat to normal brain oxygenation and function.^{34–36} The pathology of small vessel disease includes microvascular rarefaction,^{37–39} neurovascular dysfunction,⁴⁰ and disruption of the blood-brain barrier.^{41,42} It is estimated that 60–90% of AD patients have significant cerebrovascular pathology.⁴³

Hainsworth and Markus⁴⁴ identified four key clinicopathologic hallmarks of cerebral small vessel disease: small, discrete infarcts within the brain, arteriopathy of small cerebral vessels, diffuse damage to the white matter, and cognitive impairment. An Aβ-mediated positive feedback mechanism amplifies these effects of small vessel disease. With reduced capillary surface area, A^β clearance is compromised,⁴⁵ and elevated interstitial A β further depresses endothelial function.⁴⁶ Cerebral small vessel disease depresses endothelial synthesis and regulation of NO, and reduced constitutive cerebrovascular production of NO leads to further cerebral hypoperfusion.^{47,48} NO also inhibits expression of amyloid precursor protein (APP) and inactivates β -site APP-cleaving enzyme 1, which is involved in formation of $A\beta$ from APP and influences the functional status of microglia.49

The net effects of cerebral small vessel disease are impairment of cerebral oxygen and substrate delivery and diminished capacity to remove extracellular respiratory waste products, toxins, and A β . Furthermore, vascular-mediated depression of cerebral glucose and oxygen metabolism correlate positively with the severity of AD.^{50,51} Moreover, these metabolic disturbances appear to precede neurodegeneration and overt cognitive impairment in AD.⁵²⁻⁵⁴

Progressive atherosclerotic and small vessel disease chronically limit cerebral oxygen delivery while altering endothelial exchange dynamics. Thus, there is increasing appreciation that vascular therapy might play an important role in slowing or delaying progression of AD. A number of potential interventions for small cerebral vessel pathology have been identified, including neurotrophins, endothelin antagonists, NO donors, prostacyclin mimetics, phosphodiesterase inhibitors, and peroxisome proliferator-activated receptor- γ agonists.⁵⁵ However, at present there are no specific therapies approved to improve cerebrovascular perfusion beyond carotid angioplasty,⁵⁶ control of systemic arterial blood pressure⁵⁷ and blood glucose,⁵⁸ and reduction of hypercholesterolemia.⁵⁹

AD is characterized by a cerebral cholinergic deficit.¹³ While not specifically directed to the cerebral vasculature, the use of cholinesterase inhibitors to preserve cognitive function in developing AD might indirectly improve cerebral blood flow by enhancing parasympathetic-mediated dilation, or conversely, reduce flow by inactivating the α 7nicotinic acetylcholine (ACh) receptor on extrinsic sympathetic nerves and decreasing NO release.⁶⁰ In early AD, cholinesterase inhibitors have been reported to improve cerebral blood flow,^{61,62} but negative findings have also been reported.⁶³ Also, cholinesterase inhibitors increase risk of cardiovascular (bradycardia, heart block, syncope) and gastrointestinal (nausea, vomiting, diarrhea) complications.⁶⁴⁻⁶⁶ In any case, longitudinal studies of cerebral blood flow are likely confounded by the increasing severity of AD, which, by decreasing neuronal activity, decreases regional metabolism and O₂ demand (Figure 2).

Interventions to treat VRFs

Considering the paucity of direct, effective pharmacological therapies for AD, and in particular for improving cerebrovascular function, other, less direct approaches to delaying or even reversing AD merit study. Studies addressing VRF^{1,18,58} have demonstrated significant improvements in AD symptoms and progression.⁶⁷ Results of the multicenter randomized clinical trial, FINGER, showed that nutritional guidance, exercise, cognitive training, social activity, and intensive monitoring/management of metabolic/VRFs (e.g. impaired glucose tolerance, obesity, hypertension, hypercholesterolemia) reduced the risk of cognitive decline by 31% compared to a control group receiving only regular health advice.⁶⁸ In another study, patients with AD and no evidence of cerebrovascular disease were treated for VRF, including hypertension, dyslipidemia, diabetes mellitus, and smoking, for six months. The rate of cognitive decline of these treated patients was significantly slower than that of untreated patients,⁶⁹ although the treatment did not reverse AD.

There is increasing interest in physical exercise to reduce VRF and, thereby, reduce cognitive decline and dementia.^{70,71} A meta-analysis of 18 randomized trials showed that aerobic exercise training conferred significant positive benefit on cognition in patients with AD or non-AD dementia.⁷² In a recent randomized controlled trial with supervised moderate-to-high intensity exercise, patients with mild AD had reduced neuropsychiatric symptoms.⁷³ There was a tendency, also, for cognition to improve. Favorable effects of treadmill exercise were also recently reported in a rat model of experimental AD.⁷⁴ Physical exercise is well known to increase resting parasympathetic tone75 and, thus, likely improves resting cerebral blood flow. In addition, exercise has been suggested to promote angiogenesis, neurogenesis, synaptogenesis, and the synthesis of neurotransmitters in structures involved in cognition due to an increase in the liberation of neurotrophic



Figure 2 The vicious cycle of dementation and cerebral underperfusion. Decreased activity lowers energy consumption and, thus, demand for blood flow in that brain region. On the other hand, impaired local blood flow, e.g. due to cerebrovascular disease, compromises oxygen and fuel delivery and, thus, ATP production to support brain function. Severe and/or prolonged underperfusion causes neurodegeneration by the mechanisms diagramed in Figure 1. Lower right: the broken lines designate mechanisms mobilized by intermittent hypoxia training (IHT) to acutely increase metabolic vasodilation via nitric oxide (NO) formation, and the more sustained adaptive increase in microvascular density via hypoxia-inducible factor (HIF) induction of vascular endothelial growth factor (VEGF), the principal activator of angiogenesis

factors and the production of enzymatic antioxidants.^{71,76} Stimulation of cerebral microvascular NO formation is likely an important means by which exercise increases brain blood flow.⁷⁰

Chronic mental stress is a risk factor for AD.^{77–80} Since mental stress has a negative impact on cerebral vascular function,^{81–83} and, specifically, increases the risk of cerebral atherosclerosis⁸⁴ and vasospasm⁸⁵ it may be considered a VRF. Interestingly, meditation provides stress relief and improves cerebral blood flow^{86,87} and does so in subjects with memory loss.⁸⁸ In addition, meditation reduces other VRF,⁸⁹ e.g. by improving blood pressure control, decreasing insulin resistance and reducing lipid peroxidation.⁹⁰

Intermittent hypoxic training (IHT) improves cerebrovascular function and reduces VRF Characteristics of therapeutic IHT versus adverse hypoxia paradigms

The therapeutic application of moderate IHT reviewed here should not be confused with the intense, episodic intermittent hypoxia associated with sleep apnea, a pathological condition and AD risk factor^{91,92} that produces chronic hypertension and has adverse cardiovascular and cerebral effects in AD.^{15,16} In contrast, therapeutic IHT utilizes exposures to moderately severe hypoxia produced in hypobaric chambers or by inspiring normobaric atmospheres with O₂ content lowered to approximately 10%. Typical normobaric IHT protocols utilized in our laboratories for human⁹³⁻⁹⁵ and animal⁹⁶⁻¹⁰⁰ research involve 5-8 cycles/ day of inspiration of 10% O2 for 5-10 min, with intervening 4 min normoxia (21% O₂), over the course of 14–20 consecutive days. Rats and dogs subjected to these protocols showed no distress, and humans (including coauthor HFD) reported no discomfort. In other laboratories, human subjects with hypertension,¹⁰¹ ischemic heart

disease,^{102,103} and chronic obstructive pulmonary disease¹⁰⁴ have completed similar protocols with no untoward effects. When intermittent hypoxia was produced in hypobaric chambers, daily hypoxic exposures were 4 h bouts at a simulated altitude of 4000 m for 14 days. The hypoxic exposures of IHT differ from those of chronic sleep apnea, which are shorter, usually more severe, continue throughout sleep for years, and are often accompanied by the added stress of partial or complete nocturnal arousal and persistent activation of the sympathetic nervous system.¹⁰⁵

It is well known that chronic hypoxia stimulates angiogenesis in brain and other tissues.¹⁰⁶ However, chronic hypoxia is an impractical means to improve brain vascularity, and this procedure also stimulates significant pulmonary hypertension and right ventricular hypertrophy¹⁰⁷ and increased blood viscosity due to increased hematocrit.^{108,109} As might be expected, the duration of continuous hypoxia is critical for development of right ventricular hypertrophy. Recently Ma *et al.*¹¹⁰ reported that more than three weeks at 3000 m altitude was required for rats to develop pulmonary hypertension, a precursor of right ventricular hypertrophy. Since typical IHT protocols expose subjects to moderate hypoxia intermittently for less than 2 h/day over at most three weeks, the stimulus for pulmonary hypertension and right ventricular hypertrophy should be minimal. In addition, IHT stimulates vasodilatory NO production,111-113 which should delay the pulmonary hypertensive effects of hypoxia. Thus, it was not surprising that rats treated for 21 consecutive days with IHT had no right ventricular hypertrophy.¹⁰⁰

IHT improves cerebral blood flow and prevents cerebrovascular rarefaction in AD

In a rat model of experimental AD induced by bilateral injection of a fragment of A β (A β 25–35) into the basal magnocellular nuclei, cerebral blood flow in the parietal cortex

was assessed by continuous recording with a laser Doppler flowmeter. IHT normalized cerebral blood flow by reducing endothelial dysfunction caused by $A\beta^{114}$ and increased the density of blood vessels on the brain surface of rats with experimental hypertension.¹¹⁵ This finding was more recently extended to a rat model of experimental AD, in which vascular density was significantly decreased in both hippocampus and cortex by 22–25%.¹¹⁶ However, vascular density in AD rats treated with IHT did not differ from that of the respective brain areas of normal control rats. It seems likely that IHT-induced angiogenesis and the resulting prevention of brain vascular rarefaction would improve cerebral blood flow and its regional distribution.¹¹⁷

Cerebral angiogenesis is initiated in hypoxia by the transcription factor, hypoxia-inducible factor-1 (HIF-1) through activation of genes with promoter regions containing hypoxic response elements, including the vascular endothelial growth factor (VEGF) gene.^{118,119} In response to hypoxia, VEGF is concentrated primarily in the end feet of the astrocytes surrounding the capillaries¹²⁰ where it is ideally positioned to exert paracrine activation of angiogenesis. Conceivably, these mechanisms were also responsible for the cerebral angiogenesis induced by IHT,¹²¹ but this hypothesis requires further investigation. In any case, it is likely that the resulting increases in cerebral blood flow, oxygen and substrate delivery, and removal of metabolic waste products and toxins produced by IHT-induced cerebral angiogenesis would favorably impact brain function. VEGF is one of several HIF-1-responsive genes which, collectively, increase the cell's ability to withstand periods of severe hypoxia and/or ischemia. For example, HIF-1driven expression of glycolytic enzymes¹²² could increase O2-independent ATP production, and erythropoietin, long recognized for its pivotal role in hematopoiesis,¹²³ also can be synthesized within the brain, where it exerts a host of neuroprotective actions.¹²⁴

IHT improves endothelial production of NO

Normally, NO involved in cerebral blood flow regulation is produced by endothelial cells, neurons, and nitrergic perivascular nerves.¹²⁵ In addition to regulating cerebral blood flow, endothelial NO is vaso- and neuroprotective due to its antiplatelet, antithrombotic, antiproliferative, and antiatherosclerotic functions.¹²⁶ Endothelial dysfunction resulting from decreased NO production and bioavailability increases the risk of cerebral underperfusion in AD.¹²⁷⁻¹²⁹ Since aging-related neurodegeneration can be delayed by stimulation of endothelial NO synthesis,¹³⁰ interventions to prevent underproduction of endothelial NO should be valuable in prevention and treatment of AD.

Experimental AD induced by A β injection into the basal magnocellular nuclei in rats was associated with attenuated systemic production of NO as indicated by decreased plasma nitrite + nitrate, i.e. NOx.¹³¹ IHT moderately increased NO synthesis and alleviated the decrease in plasma NOx in experimental AD. The reverse was observed when NO production was limited by daily administration of N ω -nitro-*L*-arginine (*L*-NNA), a nitric oxide synthase

(NOS) inhibitor, and memory loss was exacerbated. In contrast, increasing plasma NO by administration of the NO donor, dinitrosyl iron complex, prevented the A β -induced memory impairment, similar to the effect of IHT. Experimental AD attenuated endothelium-dependent aortic vasodilation, but this effect was not altered by IHT (unpublished observation).

In rats with experimental AD, ACh was injected into a carotid artery, to assess endothelium-dependent modulation of cerebral blood flow, which was monitored continuously with a laser Doppler flowmeter.¹¹⁴ Endothelium-dependent cerebral dilation was decreased ~77% in experimental AD. Intermittent hypobaric hypoxia (4 h/day for 14 days at 4000 m simulated altitude) prevented completely this endothelial dysfunction; thus, IHT normalized ACh-induced vasodilation to that of non-AD controls.

IHT ameliorates overproduction of non-endothelial NO in AD

NO has both beneficial and detrimental actions in AD.¹³²⁻¹³⁴ Endothelial production of NO is compromised in AD, and this is one factor leading to cerebral underperfusion. On the other hand, excessive production of NO in microgia, astrocytes, and cortical neurons has been demonstrated.¹³³⁻¹³⁹ A key event that triggers NO overproduction in AD is induction of inflammation and immune responses by A β , which results in iNOS expression in microglia and astrocytes.¹³³ Another important event is glutamate excitotoxicity, which excessively increases cytosolic Ca²⁺.¹³³ The Ca²⁺ influx into postsynaptic neurons activates nNOS, which is co-localized with glutamatergic *N*-methyl-D-aspartate receptors. Together iNOS and nNOS generate toxic levels of NO.¹³³ In experimental AD, non-endothelial tissue¹⁴⁰ also contributes to excessive NO formation. Excessive NO is toxic to neurons,¹³⁹ where it condenses with superoxide to form peroxynitrite,¹¹² a powerful oxidant that irreversibly damages lipids, DNA, carbohydrates, and proteins, leading to mitochondrial dysfunction and cellular degeneration.¹⁴¹⁻¹⁴⁵ This nitrative damage occurs early in the progression of AD and may be important for transition from mild cognitive impairment to the severe impairment characteristic of AD.¹⁴⁶

To examine empirically the association of AD and NO overproduction, we measured the stable NO metabolites, nitrite and nitrate, in rat brain with experimental AD.143 The results demonstrated that $A\beta$ induced pronounced brain NO overproduction resulting from excessive expression of all three NOS isoforms. IHT completely prevented this NO overproduction in Aβ-injected rats. In these experiments, AB injections were associated with accumulation of 3-nitrotyrosine (3-NT), a product of peroxynitrite modification of protein tyrosyl residues, in cortex and in hippocampus. IHT dampened these increases in 3-NT in both brain regions. In this study, we obtained direct evidence of hypoxia-induced neuroprotection by histopathological analysis of the parietotemporal cortex. Damaged neurons were not found in control rats but multiple shrunken, hyperchromic degenerating neurons were present in the parietotemporal cortex of rats injected with AB. After IHT, neurons with these pathomorphological signs were essentially absent. Therefore, although adaptation to hypoxia *per se* moderately increases NO production,^{112,131} it prevents cytotoxic NO overproduction in experimental AD and alleviates adverse effects of AD on memory.¹⁴³

NO can be bound into *S*-nitrosothiols and dinitrosyl iron complexes for transport and intracellular storage.¹⁴⁷⁻¹⁴⁹ These NO stores are formed with any increase in NO, so by sequestering excess free NO, these stores can provide protection by buffering NO overproduction. On the other hand, when NO production is deficient, NO stores can be mobilized to provide an additional NO source.¹⁵⁰

The protective role of NO stores against NO overproduction in experimental AD was investigated in rats.¹¹⁴ The size of NO stores was evaluated from the ability of Nacetylcysteine (NAC) to release vasoactive products from intracellular NO stores, thereby increasing cerebral blood flow.^{114,148} To prevent the contribution of *de novo* synthesized NO to the cerebral vasodilatory response, NO stores were estimated in the presence of the NO synthase inhibitor, L-NNA. To reveal NO stores, NAC was injected into the carotid artery, and dilation of cerebral blood vessels was evidenced by an increase in flow measured with Doppler flowmetry. Since the size of NO stores correlates closely with NO production,¹¹¹ increased NO stores may be regarded as an indirect marker of NO overproduction. Cerebral NO stores were absent in control rats, but NAC administration revealed NO stores in both hypoxia-adapted and A\beta-treated rats.¹¹⁴ In rats injected with Aβ after IHT adaptation, NO stores were increased compared to nonadapted A\beta-treated rats. IHT increased NO storage capacity as reflected by the maximum amount of NO, which can potentially be bound to and then released from NO stores.¹¹¹ This mechanism may underlie the protection of blood vessels against subsequent NO overproduction in rats with experimental AD, as reported by Mashina et al.¹¹⁴

IHT reduces oxidative stress in AD

Oxidative stress plays a pivotal role in the pathogenesis of AD.^{151,152} Aβ deposits and activated microglia and astrocytes are major sources of reactive oxygen species (ROS) in early AD.¹⁵³ Among the numerous harmful effects of ROS overproduction, mitochondrial dysfunction contributes to energy imbalance and apoptosis of nerve cells.154-157 In addition to increased oxidative stress, AD brain is characterized by impaired antioxidant power. The ratio between reduced and oxidized glutathione, a measure of antioxidant power, is decreased in affected brain regions¹⁵⁴ and also in lymphocytes from AD patients.^{158,159} Activities of the antioxidant enzymes superoxide dismutase, glutathione peroxidase, and catalase were found to be decreased in hippocampus of rats with experimental AD.¹⁶⁰ Oxidative stress may contribute to formation of neurofibrillary tangles, too. Specifically, ONOO⁻, the product of NO-superoxide condensation, nitrotyrosinates the glycolytic enzyme triosephosphate isomerase. The modified enzyme then interacts with tau in a manner that promotes tau aggregation, forming neurofibrillary tangles.¹⁶¹

There is direct evidence that IHT potentiates antioxidant defense. Increased activities of the antioxidant enzymes, superoxide dismutase, catalase, and glutathione peroxidase, were found in erythrocytes of rats treated with IHT.¹⁶² IHT of older adult (24-month-old) rats increased the resistance of cardiac mitochondria to phenylarsineoxide-induced permeability transition pore opening and decreased myocardial accumulation of ROS.¹⁶³ Gonchar and Mankovska¹⁶⁴ and Gonchar *et al.*¹⁶⁵ demonstrated IHT of rats produced by alternating 5 min exposures to moderate, normobaric hypoxia (10% O₂) and 5 min hyperoxia (30% O₂) decreased lipid peroxidation, increased GSH/GSSG and augmented activities of the antioxidant enzymes MnSOD, glutathione peroxidase, glutathione reductase, and NADPH-generating isocitrate dehydrogenase in lung and liver mitochondria, thereby increasing mitochondrial resistance to acute, severe hypoxia. We observed a significant decrease in production of malondialdehyde, a footprint of lipid peroxidation, in the hippocampus of rats with experimental AD after adaptation to intermittent hypobaric hypoxia.^{166,167} Also, Kalachev *et al.*¹⁶⁸ reported in human subjects that IHT reduced total pro-oxidant activity and malondialdehyde concentration, and increased the activities of antioxidant enzymes in blood. In another clinical study, IHT increased antioxidant defenses in hypertensive patients with dyscirculatory encephalopathy.¹⁶⁹ Total blood oxidant activity and malondialdehyde were decreased, while superoxide dismutase, catalase, and glutathione peroxidase activities in erythrocytes were increased. Cerebral circulation and short-term memory of these patients were improved.

Regarding the antioxidant mechanism of IHT, we hypothesize that IHT causes intermittent, moderate activation of ROS formation, which occurs during the cyclic alternations of hypoxia and normoxia during the adaptation sessions. These mild, intermittent increases in ROS may induce expression of antioxidant enzymes,¹⁷⁰ most likely by activating the transcription factor Nrf2, which in turn activates expression in astrocytes of a host of antioxidant genes.¹⁷¹⁻¹⁷³ Interestingly, administration of antioxidant NAC prior to each IHT session abolished protective effects of IHT on ischemic myocardium in dogs.¹⁷⁴

The pineal hormone melatonin is an antioxidant and a free radical scavenger that is deficient in many AD patients. Melatonin also possesses antiamyloidogenic properties.¹⁷⁵ Thus, there is increasing interest in potential benefits of melatonin in AD.^{176,177} Hypoxia exposure of Wistar rats steadily increased plasma melatonin activity to a plateau at seven days.¹⁷⁸ To what extent the beneficial actions of IHT in AD are due to increased release of melatonin by the pineal gland remains to be determined.

IHT reduces VRFs

In addition to its direct effects on cerebrovascular function, IHT has been employed to effectively treat conditions associated with VRF, including systemic hypertension, atherogenic changes in the lipid profile, smoking, obesity and metabolic syndrome, ischemic cardiac disease, and psychological stress.^{101,102,179,180} While not specifically focused on VRF, a recent review by Navarrete-Opazo and Mitchell¹⁸¹ described IHT's effects on the respiratory, cardiovascular, immune, metabolic, skeletal, and nervous systems. The following discussion extends that of Navarrete-Opazo and Mitchell¹⁸¹ to more specifically address IHT's impact on VRF.

In a clinical study in patients with stage I hypertension, Lyamina *et al.*¹⁰¹ found that normobaric IHT decreased blood pressure completely to normotensive values in 92% of patients and partially in the other 8%. In 85% of the patients, the antihypertensive effect of IHT persisted for at least three months. These results supported earlier findings with hypobaric IHT.^{182,183} Those studies also reported that about 20% of the smoking subjects stopped smoking after IHT. In spontaneously hypertensive rats, IHT slowed development of hypertension and completely prevented development of vascular endothelial dysfunction.⁹⁸

Since the intermittent hypoxia of human sleep apnea is associated with hypertension, it is not surprising that some animal studies of "intermittent hypoxia" have shown elevated arterial pressure. However, those intermittent hypoxia protocols were designed to simulate the intense, episodic intermittent hypoxia of sleep apnea, clearly distinct from the IHT that we have found to be antihypertensive. For example, Perim et al.¹⁸⁴ recently concluded that intermittent hypoxia increases arterial blood pressure of rats. In their protocol, FIO2 was reduced to 6% for 30-40 s, followed by 21% O₂ for 5 min, and these hypoxia-reoxygenation cycles were repeated for 8 h/day over 10 days. Clearly the "dose" of intermittent hypoxia is critical for determining its effects on blood pressure and, most likely, other outcomes.¹⁸¹ It also should be noted that obstructive sleep apnea interrupts alveolar ventilation, producing hypercapnea, whereas IHT has the opposite effect. Studies in rats demonstrated that hypercapnia produced by addition of CO₂ to the hypoxia chamber blunted the cardioprotective effects of moderate hypoxia.¹⁸⁵

In subjects with ischemic heart disease, hypobaric IHT decreased total cholesterol by 7%, increased high-density lipoprotein by 12%, and decreased low-density lipoprotein by 13%.¹⁸⁶ These changes were greatest at three months post-treatment and persisted for six months. An atherogenic index declined by 26% immediately post-treatment and by 37% at three months post-treatment. However, conflicting data were earlier reported from studies in very young rats exposed to IHT,¹⁸⁷ yet the IHT-treated rats had lower serum low-density lipoproteins as adults.

The effect of IHT on obesity and metabolic syndrome was studied in young men aged 17–25 years.¹⁸⁸ After IHT, body weight, fat mass, and body mass index decreased significantly compared to respective values in untreated control subjects. IHT produced similar, favorable responses in postmenopausal women with metabolic syndrome,¹⁸⁹ including reduced body weight, body mass index, systolic and diastolic blood pressures, serum glucose concentration and insulin activity, and total cholesterol. IHT was found to augment the effects of exercise on obesity.¹⁹⁰

The beneficial effects of adaptation to hypoxia in ischemic cardiac disease have been recognized for many years.^{179,180,191} In 1993, Ehrenbourg and Gorbachenkov¹⁰² described IHT treatment of patients with stable angina. Remarkably, IHT decreased the frequency of angina attacks by $56.2 \pm 5.1\%$ in patients with NYHA classes I and II and by $50.7 \pm 6.1\%$ in patients with NYHA classes III and IV. IHT improved the functional class in 55% of patients. Unfortunately these studies were reported in a peerreviewed journal not readily available to western readers.

In another early study, Lyamina and Pilyavsky¹⁹¹ described favorable effects of IHT on cardiac rhythm disorders in patients with neurocirculatory dysfunction. In a more recent study, IHT was performed in patients with functional class I-IV angina who were receiving standard drug therapy for ischemic heart disease, including nitrates, β-blockers, and long-acting calcium antagonists.¹⁹² IHT abolished cardiac arrhythmias in 57% of the patients and reduced arrhythmias in the other 43%. The incidence of anginal attacks decreased by 45% after IHT. Also, this use of IHT as an adjuvant to drug therapy appeared to neither suppress myocardial contractility nor impair cardiac conductance. These clinical studies in patients are substantiated by relevant preclinical studies showing antiarrhythmic and myocardial protective effects of IHT in animals.^{96,97,100,193,194} Further information on the cardioprotective effects of IHT are available in the recent reviews by Zhang and Zhou¹⁹⁵ and by Serebrovska and Shatilo.¹⁹⁶ Various antistress effects of IHT have been described by IHT pioneer Felix Meerson.¹⁷⁹ He and his co-workers found that IHT prevented stomach ulcers induced by restraint stress and behavioral disorders in rats.¹⁹⁷ Zhu et al.198 reported that IHT produced antidepressant-like effects in rats subjected to a variety of mild stressors, e.g. restraint, forced swim in ice-cold water, and strobe light, and these effects were accompanied by hippocampal neurogenesis. In studies of alcohol withdrawal stress in rats, IHT attenuated oxidative damage to the brain and mitigated behavioral abnormalities.^{99,199} In summary, there is robust evidence that IHT can impact many of the major VRF.

While there are no reports of clinical applications of IHT in patients with AD, there has been considerable research in clinical applications of IHT for other brain pathologies, including dyscirculatory encephalopathy¹⁶⁹ and parkinsonism.²⁰⁰ In their recent review of IHT and neurologic dysfunction,²⁰¹ Gonzalez-Rothi *et al.* stated "it has become clear that brief, repeated presentations of hypoxia [i.e., acute intermittent hypoxia] can boost the efficacy of more traditional therapeutic strategies in certain cases of neurologic dysfunction." While Gonzalez-Rothi *et al.* emphasized the effects of IHT on neurorehabilitation after spinal cord injury, they noted that the fundamental principles of IHT are likely to apply to other dyskinesias.

Intermittent hypoxia training has been shown to afford neuroprotection against ischemic stroke injury. Stowe *et al.*²⁰² found that IHT of male mice provided protection against transient focal stroke that persisted for eight weeks after the IHT program. Postischemic inflammation, endothelial permeability, and leukocyte diapedesis across the blood-brain barrier were lessened. Brain infarct volumes were reduced, but cerebral blood flow during and after ischemia was not altered. These findings extended earlier reports that IHT prevented cortical infarction following cerebral ischemia-reperfusion.^{203,204} In one of these studies,²⁰³ similar reductions in brain infarct volume were afforded by IHT and ischemic preconditioning of the brain. IHT was also observed to prevent impairment of passive avoidance learning and suppression induced by severe brain hypoxia.²⁰⁵

Oligodendrocyte fusion with neurons has been observed in the poststroke rat brain.²⁰⁶ The resulting binucleated neuron would have enhanced functional capability due to its increased number of processes and synapses. This novel means of neuroregeneration may compensate to some extent for the loss of neurons and synapses during the ischemic insult. We recently found that IHT significantly increased the number of cortical binuclear neurons in normal rat brains.²⁰⁷ To what extent this mechanism contributes to the favorable effects of IHT in models of stroke remains to be determined.

While no effects of IHT in human stroke have been reported, it is interesting to note that ischemic stroke patients who had previously had spontaneous, transient cerebral ischemic attacks had more favorable neurocognitive outcomes than patients without previous transient attacks.²⁰⁸ Prior transient ischemic attacks of approximately 10 min duration appeared to provide the optimum protective effect.

Intermittent hypoxia training also had positive effects in rats of the KM strain, which are predisposed to audiogenic epilepsy induced by loud noises.209 Seizures were prevented and the associated subdural hemorrhage was significantly less in IHT than in non-IHT rats. This antiepileptic effect of IHT was confirmed in a recent study by Zhen et al.,²¹⁰ where IHT reduced the frequency and severity of seizures in rats with pilocarpine-induced epilepsy. This protection was associated with suppression of intracellular calcium overload and inhibition of neuronal apoptosis in the hippocampus. The antiepileptic effect of IHT may also involve upregulation of monocarboxylate transporter 4 expression in the plasma membranes of rat hippocampal astrocytes,²¹¹ an adaptation that could increase the astrocyte's capacity to supply lactate as energy substrate to the adjacent neurons.²¹² In patients with encephalopathy,¹⁶⁹ IHT reduced the elevated peak systolic, peak diastolic, and mean blood flow velocities in the middle cerebral artery. Importantly, IHT alleviated headache and improved sleep and memory function of the patients.

Conclusions and perspectives

The potential limitations of IHT intervention for AD must be acknowledged. As reviewed here, many AD patients have advanced cerebrovascular disease, and while IHT may be safe and readily tolerated by persons with intact cerebral circulations, it is not certain if even brief periods of moderate hypoxia would be without untoward effects in senescent patients with low cerebral blood flow or carotid artery stenosis. However, IHT can be used to modify VRF and therefore may be efficacious in patients who have already had predisposing conditions, such as stroke, transient ischemic attacks, diabetic or dyscirculatory encephalopathy, and post-traumatic stress. Although beyond the scope of this review, it also should be noted that IHT is not only vasoprotective but potentially neuroprotective in AD, as it prevents neuronal degeneration and stimulates neurogenesis and neuroregeneration.^{166,167,207}

Another important concern is whether or not IHT would be effective in patients with clinically significant dementia. Because the complex, insidious pathogenesis of AD may begin decades before neurocognitive impairment is manifest, the disease is already far advanced by the time overt dementia is detected, and the opportunity to interrupt its progression may have been missed. On the other hand, the recent identification of the APO-E4*ɛ* gene as a major risk factor for AD may afford opportunities for effective, early intervention with IHT. As noted above, 40-50% of AD patients carry the APO-E4*ε* allele.¹³ Timely genetic testing could afford early identification of APO-E4ε, when IHT therapy could be initiated in time to slow progression or even prevent development of AD. Such clinical application of IHT as an early intervention for APO-E4ɛ-associated AD merits investigation.

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REFERENCES

- de la Torre JC. Vascular risk factors: a ticking time bomb to Alzheimer's disease. Am J Alzheimers Dis Other Demen 2013;28:551–9
- Reitz C, Mayeux R. Alzheimer disease: epidemiology, diagnostic criteria, risk factors and biomarkers. *Biochem Pharmacol* 2014;88:640–51
- Cummings JL, Vinters HV, Cole GM, Khachaturian ZS. Alzheimer's disease: etiologies, pathophysiology, cognitive reserve, and treatment opportunities. *Neurology* 1998;51:S2–17
- Jamasbi E, Wade JD, Separovic F, Hossain MA. Amyloid beta (Aβ) peptide and factors that play important roles in Alzheimer's disease. *Curr Med Chem* 2016;23:884–92
- Khan SS, Bloom GS. Tau: the center of a signaling nexus in Alzheimer's disease. Front Neurosci 2016;10:31
- 6. Sun X, Chen WD, Wang YD. β-Amyloid: the key peptide in the pathogenesis of Alzheimer's disease. *Front Pharmacol* 2015;6:221
- Jin K, Peel AL, Mao XO, Xie L, Cottrell BA, Henshall DC, Greenberg DA. Increased hippocampal neurogenesis in Alzheimer's disease. *Proc Natl Acad Sci USA* 2004;101:343–7
- 8. Hebert LE, Scherr PA, Bienias JL, Bennett DA, Evans DA. Alzheimer disease in the US population. *Arch Neurol* 2003;**60**:1119–22
- Pahnke J, Walker LC, Scheffler K, Krohn M. Alzheimer's disease and blood-brain barrier function – why have anti-β-amyloid therapies failed to prevent dementia progression? *Neurosci Behav Rev* 2009;**33**:1099–108

 Ballard C, Gauthier S, Corbett A, Brayne C, Aarsland D, Jones E. Alzheimer's disease. *Lancet* 2011;377:1019–31

- Honjo K, Black SE, Verhoeff NPLG. Alzheimer's disease, cerebrovascular disease, and the β-amyloid cascade. *Can J Neurol Sci* 2012;39:712–28
- Amemori T, Jendelova P, Ruzicka J, Urdzikova LM, Sykova E. Alzheimer's disease: mechanism and approach to cell therapy. *Int J Mol Sci* 2015;16:26417–51
- Gandy S, DeKosky ST. Toward the treatment and prevention of Alzheimer's disease: rational strategies and recent progress. Annu Rev Med 2013;64:367–83
- Feldstein CA. Association between chronic blood pressure changes and development of Alzheimer's disease. J Alzheimer's Dis 2012;32:753–63
- Bu XL, Liu YH, Wang QH, Jiao SS, Zeo XQ, Gao D, Chen JC, Wang YJ. Serum amyloid-beta levels are increased in patients with obstructive sleep apnea syndrome. *Sci Res* 2015;5:13917
- Daulatzai MA. Evidence of neurodegeneration in obstructive sleep apnea: relationship between obstructive sleep apnea and cognitive dysfunction in the elderly. J Neurosci Res 2015;93:1778–94
- Kalaria RN, Akinyemi R, Ihara M. Does vascular pathology contribute to Alzheimer changes? J Neurol Sci 2012;322:141–7
- Akinyemi RO, Mukaetova-Ladinska EB, Attems J, Ihara M, Kalaria RN. Vascular risk factors and neurodegeneration in ageing related dementias: Alzheimer's disease and vascular dementia. *Curr Alzheimer Res* 2013;10:642–53
- Barclay L, Zemcov A, Blass JP, McDowell F. Rates of decrease of cerebral blood flow in progressive dementias. *Neurology* 1984;34:1555–60
- Hanyu H, Sato T, Hirao K, Kanetaka H, Iwamoto T, Koizumi K. The progression of cognitive deterioration and regional cerebral blood flow patterns in Alzheimer's disease: a longitudinal SPECT study. J Neurol Sci 2010;290:96–101
- Rius-Pérez S, Tormos AM, Pérez S, Taléns-Visconti R. Vascular pathology: cause or effect in Alzheimer disease? *Neurologia* 2015;pii:S0213-4853(15)00185-1
- Bai XJ, Iwamoto T, Williams AG Jr, Fan WL, Downey HF. Coronary pressure-flow autoregulation protects myocardium from pressureinduced changes in oxygen consumption. *Am J Physiol Heart Circ Physiol* 1994;**266**:H2359-68
- Ito H, Kanno I, Fukuda H. Human cerebral circulation: positron emission tomography studies. *Ann Nucl Med* 2005;19:65–74
- Siesjö BK. Cerebral circulation and metabolism. J Neurosurg 1984;60:883–908
- Armstead WM. Role of opioids in the physiologic and pathophysiologic control of the cerebral circulation. Proc Soc Exp Biol Med 1997;214:210–21
- Bor-Seng-Shu E, Kita WS, Figueiredo EG, Paiva WS, Fonoff ET, Teixeira MJ, Panerai RB. Cerebral hemodynamics: concepts of clinical importance. *Arq Neuropsiquiatr* 2012;70:352–6
- Park SS, Hong M, Ha Y, Sim J, Jhon GJ, Lee Y, Suh M. The real-time in vivo electrochemical measurement of nitric oxide and carbon monoxide release upon direct epidural electrical stimulation of the rat neocortex. *Analyst* 2015;**140**:3415–21
- Kontos HA, Wei EP. Oxygen-dependent mechanisms in cerebral autoregulation. Ann Biomed Eng 1985;13:329–34
- Niwa K, Kazama K, Younkin L, Younkin SG, Carlson GA, Iadecola C. Cerebrovascular autoregulation is profoundly impaired in mice overexpressing amyloid precursor protein. *Am J Physiol Heart Circ Physiol* 2002;283:H315–23
- Zhao Y, Gong CX. From chronic cerebral hypoperfusion to Alzheimerlike brain pathology and neurodegeneration. *Cell Mol Neurobiol* 2015;35:101-10
- Saura H, Ogasawara K, Beppu T, Yoshida K, Kobayashi M, Yoshida K, Terasaki K, Takai Y, Ogawa A. Hypoxic viable tissue in human chronic cerebral ischemia because of unilateral major cerebral artery stenoocclusive disease. *Stroke* 2015;46:1250–6
- 32. Sagare AP, Bell RD, Zlokovic BV. Neurovascular defects and faulty amyloid β-vascular peptide clearance in Alzheimer disease. J Alzheimers Dis 2013;33:S87–100

- Weller RO, Djuanda E, Yow HY, Carare RO. Lymphatic drainage of the brain and the pathophysiology of neurological disease. *Acta Neuropathol* 2009;117:1–14
- Hennerici M. Vascular dementia: a changing concept. Arzneimittelforschung 1995;45:366–70
- Huynh TJ, Murphy B, Pettersen JA, Tu H, Sahlas DJ, Zhang L, Symons SP, Black S, Lee TY, Aviv RI. CT perfusion quantification of small-vessel ischemic severity. *Am J Neuroradiol* 2008;29:1831–6
- 36. Østergaard L, Engedal TS, Moreton F, Hansen MB, Wardlaw JM, Dalkara T, Markus HS, Muir KW. Cerebral small vessel disease: capillary pathways to stroke and cognitive decline. J Cereb Blood Flow Metab 2016;36:302–25
- Englund E. Neuropathy of white matter changes in Alzheimer's disease and vascular dementia. *Dement Geriatr Cogn Disord* 1998;9:6–12
- 38. Joutel A, Monet-Leprêtre M, Gosele C, Baron-Menguy C, Hammes A, Schmidt S, Lemaire-Carrette B, Domenga V, Schedl A, Lacombe P, Hubner N. Cerebrovascular dysfunction and microcirculation rarefaction precede white matter lesions in a mouse genetic model of cerebral ischemic small vessel disease. J Clin Invest 2010;120:433–45
- Brown WR, Thore CR. Review: cerebral microvascular pathology in ageing and neurodegeneration. *Neuropathol Appl Neurobiol* 2011;37:56–74
- Nelson AR, Sweeney MD, Sagare AP, Zlokovic BV. Neurovascular dysfunction and neurodegeneration in dementia and Alzheimer's disease. *Biochim Biophys Acta* 2015;1862:887–900
- Starr JM, Wardlaw J, Ferguson K, MacLullich A, Deary IJ, Marshall I. Increased blood-brain barrier permeability in type II diabetes demonstrated by gadolinium magnetic resonance imaging. J Neurol Neurosurg Psychiatry 2003;74:70–6
- Ueno M, Chiba Y, Matsumoto K, Murakami R, Fujihara R, Kawauchi M, Miyanaka H, Nakagawa T. Blood-brain barrier damage in vascular dementia. *Neuropathology* 2016;36:115–24
- Kalaria RN. The role of cerebral ischemia in Alzheimer's disease. Neurobiol Aging 2000;21:321–30
- 44. Hainsworth AH, Markus HS. Do in vivo experimental models reflect human cerebral small vessel disease? A systematic review. J Cereb Blood Flow Metab 2008;28:1877–91
- Zlokovic BV. Neurovascular pathways to neurodegeneration in Alzheimer's disease and other disorders. Nat Rev Neurosci 2011;12:723–38
- Gentile MT, Vecchione C, Maffei A, Aretini A, Marino G, Poulet R, Capobianco L, Selvetella G, Lembo G. Mechanisms of soluble betaamyloid impairment of endothelial function. *J Biol Chem* 2004;**279**:48135–42
- de la Torre JC, Stefano GB. Evidence that Alzheimer's disease is a microvascular disorder: the role of constitutive nitric oxide. *Brain Res Rev* 2000;34:119–36
- Toda N, Okamura T. Cerebral blood flow regulation by nitric oxide in Alzheimer's disease. J Alzheimers Dis 2012;32:569–78
- Katusic ZS, Austin SA. Endothelial nitric oxide: protector of a healthy mind. *Eur Heart J* 2014;35:888–94
- Ferreira IL, Resende R, Ferreiro E, Rego AC, Pereira CF. Multiple defects in energy metabolism in Alzheimer's disease. *Curr Drug Targets* 2010;11:1193–1206
- Meier-Ruge W, Bertoni-Freddari C. The significance of glucose turnover in the brain in the pathogenic mechanisms of Alzheimer's disease. *Rev Neurosci* 1996;7:1–19
- 52. Duara R, Barker WW, Chang J, Yoshii F, Loewenstein DA, Pascal S. Viability of neocortical function shown in behavioral activation state PET studies in Alzheimer disease. J Cereb Blood Flow Metab 1992;12:927–34
- Mosconi L. Glucose metabolism in normal aging and Alzheimer's disease: methodological and physiological considerations for PET studies. *Clin Transl Imaging* 2013;1
- 54. Sato N, Morishita R. The roles of lipid and glucose metabolism in modulation of β-amyloid, tau, and neurodegeneration in the pathogenesis of Alzheimer disease. *Front Aging Neurosci* 2015;7:199

- Bath PM, Wardlaw JM. Pharmacological treatment and prevention of cerebral small vessel disease: a review of potential interventions. *Int J Stroke* 2015;10:469–78
- 56. Antonopoulos CN, Kakisis JD, Sfyroeras GS, Moulakakis KG, Kallinis A, Giannakopoulos T, Liapis CD. The impact of carotid artery stenting on cognitive function in patients with extracranial carotid artery stenosis. *Ann Vasc Surg* 2015;**29**:457–69
- Hughes TM, Sink KM. Hypertension and its role in cognitive function: current evidence and challenges for the future. *Am J Hypertens* 2015;29:149–57
- 58. de la Torre JC. Cerebral hemodynamics and vascular risk factors: setting the stage for Alzheimer's disease. *J Alzheimers Dis* 2012;**32**:553–67
- Dias HK, Brown CL, Polidori MC, Lip GY, Griffiths HR. LDL-lipids from patients with hypercholesterolaemia and Alzheimer's disease are inflammatory to microvascular endothelial cells: mitigation by statin intervention. *Clin Sci (Lond)* 2015;**129**:1195–206
- 60. Mozayan M, Chen MF, Si M, Chen PY, Premkumar LS, Lee TJ. Cholinesterase inhibitor blockade and its prevention by statins of sympathetic alpha7-nAChR-mediated cerebral nitrergic neurogenic vasodilation. J Cereb Blood Flow Metab 2006;26:1562–76
- Chaudhary S, Scouten A, Schwindt G, Janik R, Lee W, Sled JG, Black SE, Stefanovic B. Hemodynamic effects of cholinesterase inhibition in mild Alzheimer's disease. J Magn Reson Imaging 2013;38:26–35
- 62. Li W, Antuono PG, Xie C, Chen G, Jones JL, Ward BD, Franczak MB, Goveas JS, Li SJ. Changes in regional cerebral blood flow and functional connectivity in the cholinergic pathway associated with cognitive performance in subjects with mild Alzheimer's disease after 12-week donepezil treatment. *Neuroimage* 2012;60:1083–91
- 63. Kimura N, Kumamoto T, Masuda T, Hanaoka T, Okazaki T, Arakawa R. Evaluation of the regional cerebral blood flow changes during longterm donepezil therapy in patients with Alzheimer's disease using 3DSRT. J Neuroimaging 2012;22:299–304
- 64. Birks J. Cholinesterase inhibitors for Alzheimers' disease. Cochrane Database Syst Rev 2006;25:CD005593
- Parke-Wyllie LY, Mamdani MM, Li P, Gill SS, Laupacis A, Juurlink DN. Cholinesterase inhibitors and hospitalizations for bradycardia. *PLoS Med* 2009;6:e1000157
- Howes LG. Cardiovascular effects of drugs used to treat Alzheimer's disease. Drug Saf 2014;37:391–5
- 67. Sindi S, Mangialasche F, Kivipelto M. Advances in the prevention of Alzheimer's Disease. *F1000Prime Rep* 2015;**7**:50
- 68. Ngandu T, Lehtisalo J, Solomon A, Levälahti E, Ahtiluoto S, Antikainen R, Bäckman L, Hanninen T, Jula A, Laatikainen T, Lindström J, Mangialasche F, Paajanen T, Pajala S, Peltonen M, Rauramaa R, Stigsdotter-Neely A, Strandberg T, Tuomilehto J, Soininen H, Kivipelto M. A 2 year multidomain intervention of diet, exercise, cognitive training, and vascular risk monitoring versus control to prevent cognitive decline in at-risk elderly people (FINGER): a randomised controlled trial. *Lancet* 2015;385:2255–63
- Deschaintre Y, Richard F, Leys D, Pasquier F. Treatment of vascular risk factors is associated with slower decline in Alzheimer disease. *Neurology* 2009;73:674–80
- Lange-Asschenfeldt C, Kojda G. Alzheimer's disease, cerebrovascular dysfunction and the benefits of exercise: from vessels to neurons. *Exp Gerontol* 2008;43:499–504
- Paillard T. Preventive effects of regular physical exercise against cognitive decline and the risk of dementia with age advancement. *Sports Med Open* 2015;1:20
- 72. Groot C, Hooghiemstra AM, Raijmakers PG, van Berckel BN, Scheltens P, Scherder EJ, van der Flier WM, Ossenkoppele R. The effect of physical activity on cognitive function in patients with dementia: a meta-analysis of randomized control trials. *Ageing Res Rev* 2016;25:13–23
- 73. Hoffmann K, Sobol NA, Frederiksen KS, Beyer N, Vogel A, Vestergaard K, Brændgaard H, Gottrup H, Lolk A, Wermuth L, Jacobsen S, Laugesen LP, Gergelyffy RG, Høgh P, Bjerregaard E, Andersen BB, Siersma V, Johannsen P, Cotman CW, Waldemar G, Hasselbalch SG. Moderate-to-high intensity physical exercise in

patients with Alzheimer's disease: a randomized controlled trial. *J Alzheimers Dis* 2015;**50**:443–53

- 74. Dao AT, Zagaar MA, Levine AT, Alkadhi KA. Comparison of the effect of exercise on late-phase LTP of the dentate gyrus and CA1 of Alzheimer's disease model. *Mol Neurobiol*, Epub ahead of print 12 December 2015. DOI: 10.1007/s12035-015-9612-5
- Gupt AM, Kumar M, Sharma RK, Misra R, Gupt A. Effect of moderate aerobic exercise training on autonomic functions and its correlation with the antioxidant status. *Indian J Physiol Pharmacol* 2015;59:162–9
- Paillard T, Rolland Y, de Souto Barreto P. Protective effects of physical exercise in Alzheimer's disease and Parkinson's disease: a narrative review. J Clin Neurol 2015;11:212–9
- 77. Marcello E, Gardoni F, Di Luca M. Alzheimer's disease and modern lifestyle: what is the role of stress? *J Neurochem* 2015;**134**:795–8
- Baglietto-Vargas D, Chen Y, Suh D, Ager RR, Rodriguez-Ortiz CJ, Medeiros R, Myczek K, Green KN, Baram TZ, LaFerla FM. Short-term modern life-like stress exacerbates Aβ-pathology and synapse loss in 3xTg-AD mice. J Neurochem 2015;134:915–26
- 79. Justice NJ, Huang L, Tian JB, Cole A, Pruski M, Hunt AJ Jr, Flores R, Zhu MX, Arenkiel BR, Zheng H. Posttraumatic stress disorder-like induction elevates β-amyloid levels, which directly activates corticotropin-releasing factor neurons to exacerbate stress responses. *J Neurosci* 2015;**35**:2612–23
- Greenberg MS, Tanev K, Marin MF, Pitman RK. Stress, PTSD, and dementia. Alzheimers Dement 2014;10:S155–65
- Endo Y, Nishimura JI, Kobayashi S, Kimura F. Chronic stress exposure influences local cerebral blood flow in the rat hippocampus. *Neuroscience* 1999;93:551–5
- Konorova IL, Matsievskii DD, Timkina MI. Hemodynamic mechanisms of negative impact of emotional stress on the development of cerebral ischemia in stressor load-sensitive rats. *Patol Fiziol Eksp Ter* 2009;3:10–5 [Russian]
- Balkaya M, Prinz V, Custodis F, Gertz K, Kronenberg G, Kroeber J, Fink K, Plehm R, Gass P, Laufs U, Endres M. Stress worsens endothelial function and ischemic stroke via glucocorticoids. *Stroke* 2011;42:3258–64
- Loganovsky KN, Zdanevich NA. Cerebral basis of posttraumatic stress disorder following the Chernobyl disaster. CNS Spectr 2013;18:95–102
- Dikanović M, Demarin V, Kadojić D, Kadojić M, Trkanjec Z, Titlić M, Bitunjac M, Soldo-Butković S. Effect of elevated catecholamine levels on cerebral hemodynamics in patients with chronic post-traumatic stress disorder. *Coll Antropol* 2011;35:471–5
- Newberg AB, Wintering N, Waldman MR, Amen D, Khalsa DS, Alavi A. Cerebral blood flow differences between long-term meditators and nonmeditators. *Conscious Cogn* 2010;19:899–905
- Tang YY, Lu Q, Feng H, Tang R, Posner MI. Short-term meditation increases blood flow in anterior cingulate cortex and insula. *Front Psychol* 2015;6:212
- Newberg AB, Wintering N, Khalsa DS, Roggenkamp H, Waldman RW. Meditation effects on cognitive function and cerebral blood flow in subjects with memory loss: a preliminary study. J Alzheimers Disease 2010;20:517–26
- 89. Khalsa DS. Stress, meditation, and Alzheimer's disease prevention: where the evidence stands. *J Alzheimers Dis* 2015;**48**:1–12
- Koike MK, Cardoso R. Meditation can produce beneficial effects to prevent cardiovascular disease. *Horm Mol Biol Clin Investig* 2014;18:137–43
- Pan W, Kastin AJ. Can sleep apnea cause Alzheimer's disease? Neurosci Biobehav Rev 2014;47:656–69
- Buratti L, Viticchi G, Falsetti L, Cagnetti C, Luzzi S, Bartolini M, Provinciali L, Silvestrini M. Vascular impairment in Alzheimer's disease: the role of obstructive sleep apnea. J Alzheimers Dis 2014;38:445–53
- Zhang P, Downey HF, Shi X. Acute intermittent hypoxia exposures enhance arterial oxygen delivery. *Exp Biol Med* 2010;235:1134–41
- Zhang P, Downey HF, Chen S, Shi X. Two-week normobaric intermittent hypoxia exposures enhance oxyhemoglobin equilibrium and cardiac responses during hypoxemia. *Am J Physiol Regul Integr Comp Physiol* 2014;**307**:R721–30

95. Zhang P, Shi X, Downey HF. Two-week normobaric intermittenthypoxic exposures stabilize cerebral perfusion during hypocapnia and hypercapnia. *Exp Biol Med* 2015;**240**:961–8

- Zong P, Setty S, Sun W, Martinez R, Tune JD, Ehrenburg IV, Tkatchouk EN, Mallet RT, Downey HF. Intermittent hypoxic training protects canine myocardium from infarction. *Exp Biol Med* 2004;**229**:806–12
- Mallet RT, Ryou MG, Williams AG Jr, Howard L, Downey HF. β₁-Adrenergic receptor antagonism abrogates cardioprotective effects of intermittent hypoxia. *Basic Res Cardiol* 2006;**101**:436–46
- Manukhina EB, Jasti D, Vanin AF, Downey HF. Intermittent hypoxia conditioning prevents endothelial dysfunction and improves nitric oxide storage in spontaneously hypertensive rats. *Exp Biol Med* 2011;**236**:867–73
- Ju X, Mallet RT, Downey HF, Metzger DB, Jung ME. Intermittent hypoxia conditioning protects mitochondrial cytochrome c oxidase of rat cerebellum from ethanol withdrawal stress. J Appl Physiol 2012:112:1706–14
- 100. Manukhina EB, Belkina LM, Terekhina OL, Abramochkin DV, Smirnova EA, Budanova OP, Mallet RT, Downey HF. Normobaric, intermittent hypoxia conditioning is cardio- and vasoprotective in rats. *Exp Biol Med* 2013;238:1413–20
- 101. Lyamina NP, Lyamina SV, Senchiknin VN, Mallet RT, Downey HF, Manukhina EB. Normobaric hypoxia conditioning reduces blood pressure and normalizes nitric oxide synthesis in patients with arterial hypertension. J Hypertens 2011;29:2265–72
- Ehrenbourg IV, Gorbachenkov AA. Interval hypoxic training: development of antianginal effects in patients with different functional classes of stable angina. *Hyp Med J* 1993;1:12–5
- 103. Burtscher M, Haider T, Domej W, Linser T, Gatterer H, Faulhaber M, Pocecco E, Ehrenburg I, Tkatchuk E, Koch R, Bernardi L. Intermittent hypoxia increases exercise tolerance in patients at risk for or with mild COPD. *Respir Physiol Neurobiol* 2009;**165**:97–103
- 104. Faulhaber M, Gatterer H, Haider T, Linser T, Netzer N, Burtscher M. Heart rate and blood pressure responses during hypoxic cycles of a 3week intermittent hypoxia breathing program in patients at risk for or with mild COPD. Int J Chron Obstruct Pulmon Dis 2015;10:339–45
- 105. Serebrovskaya TV, Manukhina EB, Smith ML, Downey HF, Mallet RT. Intermittent hypoxia: cause of or therapy for systemic hypotension? *Exp Biol Med* 2008;233:627–50
- Harik SI, Hritz MA, LaManna JC. Hypoxia-induced brain angiogenesis in the adult rat. J Physiol 1995;485:525–30
- 107. Kentera D, Zdravković M. In vitro contractility of the hypertrophied right ventricle of rats with pulmonary hypertension due to confinement in "hypoxic cages". *Pflügers Arch* 1976;**363**:235–8
- Schmidt W, Spielvogel H, Eckardt KU, Quintela A, Peñaloza R. Effects of chronic hypoxia and exercise on plasma erythropoietin in highaltitude residents. J Appl Physiol 1993;74:1874–8
- 109. Heinicke K, Prommer N, Cajigal J, Viola T, Behn C, Schmidt W. Longterm exposure to intermittent hypoxia results in increased hemoglobin mass, reduced plasma volume, and elevated erythropoietin plasma levels in man. *Eur J Appl Physiol* 2003;88:535–43
- 110. Ma TT, Wang Y, Zhou XL, Jiang H, Guo R, Jia LN, Chang H, Gao Y, Yao XY, Gao ZM, Pan L. Research on rat models of hypobaric hypoxiainduced pulmonary hypertension. *Eur Rev Med Pharmacol Sci* 2015;19:3723–30
- Manukhina EB, Malyshev IYu, Smirin BV, Mashina SYu, Saltykova VA, Vanin AF. Production and storage of nitric oxide in adaptation to hypoxia. *Nitric Oxide* 1999;3:393–401
- 112. Manukhina EB, Downey HF, Mallet RT. Role of nitric oxide in cardiovascular adaptation to intermittent hypoxia. *Exp Biol Med* 2006;**231**:343–65
- 113. Ryou MG, Sun J, Oguayo KN, Manukhina EB, Downey HF, Mallet RT. Hypoxic conditioning suppresses nitric oxide production upon myocardial reperfusion. *Exp Biol Med* 2008;233:766–74
- 114. Mashina SY, Aleksandrin VV, Goryacheva AV, Vlasova MA, Vanin AF, Malyshev IY, Manukhina EB. Adaptation to hypoxia prevents disturbances in cerebral blood flow during neurodegenerative process. *Bull Exp Biol Med* 2006;**142**:169–72

- 115. Manukhina EB, Sokolova IA, Rodionov IM. Changes in density of the vascular network on brain surface in rats in experiments hypertension and adaptation to high altitude. *Kardiologiya* 1982;10:1982[Russian]
- 116. Goryacheva AV, Barskov IV, Viktorov IV, Downey HF, Manukhina EB. Adaptation to intermittent hypoxia prevents rarefaction of the brain vascular net in rats with experimental Alzheimer's disease. *FASEB J* 2011;25:669.3
- 117. Ambrose CT. A therapeutic approach for senile dementias: neuroangiogenesis. J Alzheimers Dis 2015;43:1–17
- Chávez JC, Agani F, Pichiule P, LaManna JC. Expression of hypoxic inducible factor 1alpha in the brain of rats during chronic hypoxia. *J. Appl Physiol* 2000;89:1937–42
- 119. Pichiule P, LaManna JC. Angiopoietin-2 and rat brain capillary remodeling during adaptation and deadaptation to prolonged mild hypoxia. J Appl Physiol 2002;93:1131–9
- 120. LaManna JC, Chavez JC, Pichiule P. Structural and functional adaptation to hypoxia in the rat brain. *J Exp Biol* 2004;**207**:3163–9
- 121. El'chaninova SA, Koreniak NA, Pavlovskaia LI, Smagina IV, Makarenko VV. The effect of interval hypoxic hypoxia on the vascular endothelial growth factor and basic fibroblast growth factor concentrations in the peripheral blood. *Fiziol Cheloveka* 2004;**30**:93–5 [Russian]
- 122. Kunze R, Zhou W, Veltkamp R, Wielockx B, Breier G, Marti HH. Neuron-specific prolyl-4-hydroxylase domain 2 knockout reduces brain injury after transient cerebral ischemia. *Stroke* 2012;**43**:2748–56
- 123. Fisher JW. Erythropoietin: physiology and pharmacology update. *Exp Biol Med* 2003;228:1–14
- 124. Nguyen AQ, Cherry BH, Scott GF, Ryou MG, Mallet RT. Erythropoietin: powerful protection of ischemic and post-ischemic brain. *Exp Biol Med* 2014;239:1461–75
- 125. Atochin DN, Huang PL. Role of endothelial nitric oxide in cerebrovascular regulation. *Curr Pharm Biotechnol* 2011;**12**:1334–42
- Toda N, Ayajiki K, Okamura T. Cerebral blood flow regulation by nitric oxide in neurological disorders. *Can J Physiol Pharmacol* 2009;87:581–94
- 127. Austin SA, Santhanam AV, Hinton DJ, Choi DS, Katusic ZS. Endothelial nitric oxide deficiency promotes Alzheimer's disease pathology. J Neurochem 2013;127:691–700
- 128. Tan XL, Xue YQ, Ma T, Wang X, Li JJ, Lan L, Malik KU, McDonald MP, Dopico AM, Liao FF. Partial eNOS deficiency causes spontaneous thrombotic cerebral infarction, amyloid angiopathy and cognitive impairment. *Mol Neurodegener* 2015;**10**:24
- 129. Lamoke F, Mazzone V, Persichini T, Maraschi A, Harris MB, Venema RC, Colasanti M, Gliozzi M, Muscoli C, Bartoli M, Mollace V. Amyloid β peptide-induced inhibition of endothelial nitric oxide production involves oxidative stress-mediated constitutive eNOS/ HSP90 interaction and disruption of agonist-mediated Akt activation. *J Neuroinflammation* 2015;**12**:84
- McCarty MF. Vascular nitric oxide may lessen Alzheimer's risk. Med Hypotheses 1998;51:465–76
- 131. Manukhina EB, Pshennikova MG, Goryacheva AV, Khomenko IP, Mashina SY, Pokidyshev DA, Malyshev IY. Role of nitric oxide in prevention of cognitive disorders in neurodegenerative brain injuries in rats. *Bull Exp Biol Med* 2008;**146**:391–95
- 132. Manukhina EB, Downey HF, Mallet RT, Malyshev IYu. Protective and damaging effects of periodic cerebral hypoxia: the role of nitric oxide. *Vestn Ross Akad Med Nauk* 2007;2:25–33 [Russian]
- Balez R, Ooi L. Getting to NO Alzheimer's Disease: neuroprotection versus neurotoxicity mediated by nitric oxide. Oxid Med Cell Longev 2016;2016:3806157
- Hannibal L. Nitric oxide homeostasis in neurodegenerative diseases. Curr Alzheimer Res 2016;13:135-49
- 135. Ishii K, Muelhauser F, Liebl U, Picard M, Kühl S, Penke B, Bayer T, Wiessler M, Hennerici M, Beyreuther K, Hartmann T, Fassbender K. Subacute NO generation induced by Alzheimer's β-amyloid in the living brain: reversal by inhibition of the inducible NO synthase. *FASEB J* 2000;14:1485–9
- 136. Lüth HJ, Holzer M, Gertz HJ, Arendt TH. Aberrant expression of nNOS in pyramidal neurons in Alzheimer's disease is highly colocalized with p21ras and p16INK4a. *Brain Res* 2000;852:45–55

- 137. Lüth HJ, Holzer M, Gärtner U, Staufenbiel M, Arendt T. Expression of endothelial and inducible NOS-isoforms is increased in Alzheimer's disease, in APP23 transgenic mice and after experimental brain lesion in rat: evidence for an induction by amyloid pathology. *Brain Res* 2001;**913**:57–67
- 138. Fernández-Vizarra P, Fernández AP, Castro-Blanco S, Encinas JM, Serrano J, Bentura ML, Muñoz P, Martínez-Murillo R, Rodrigo J. Expression of nitric oxide system in clinically evaluated cases of Alzheimer's disease. *Neurobiol Dis* 2004;**15**:287–305
- Brown GC, Vilalta A. How microglia kill neurons. Brain Res 2015;1628:288–97
- 140. Lüth HJ, Münch G, Arendt T. Aberrant expression of NOS isoforms in Alzheimer's disease is structurally related to nitrotyrosine formation. *Brain Res* 2002;953:135–43
- Chung KK, David KK. Emerging roles of nitric oxide in neurodegeneration. Nitric Oxide 2010;22:290-5
- García-Escudero V, Martín-Maestro P, Perry G, Avila J. Deconstructing mitochondria dysfunction in Alzheimer disease. Oxid Med Cell Longev 2013;2013:162152
- 143. Goryacheva AV, Kruglov SV, Pshennikova MG, Smirin BV, Malyshev IY, Barskov IV, Viktorov IV, Downey HF, Manukhina EB. Adaptation to intermittent hypoxia restricts nitric oxide overproduction and prevents beta-amyloid toxicity in rat brain. *Nitric Oxide* 2010;23:289–99
- 144. Drechsel DA, Estévez AG, Barbeito L, Beckman JS. Nitric oxidemediated oxidative damage and the progressive demise of motor neurons in ALS. *Neurotox Res* 2012;**22**:251–64
- 145. Radi R. Peroxynitrite, a stealthy biological oxidant. J Biol Chem 2013;288:26464–72
- 146. Butterfield DA, Reed TT, Perluigi M, De Marco C, Coccia R, Keller JN, Markesbery WR, Sultana R. Elevated levels of 3-nitrotyrosine in brain from subjects with amnestic mild cognitive impairment: implications for the role of nitration in the progression of Alzheimer's disease. *Brain Res* 2007;**1148**:243–8
- 147. Mashina SY, Vanin AF, Serezhenkov VA, Kubrina LN, Malenkova IV, Malyshev IY, Manukhina EB. Detection and evaluation of NO stores in awake rats. *Bull Exp Biol Med* 2003;**136**:26–9
- 148. Vlasova MA, Vanin AF, Muller B, Smirin BV, Malyshev IY, Manukhina EB. Detection and description of various stores of nitric oxide store in vascular wall. *Bull Exp Biol Med* 2003;**136**:226–30
- 149. Manukhina EB, Vanin AF, Malyshev IYu, Mallet RT, Downey HF. Intermittent hypoxia-induced cardio- and vasoprotection: role of NO stores. In: Xi L, Serebrovskaya TV (eds). *Intermittent hypoxia: from* molecular mechanisms to clinical applications. New York: Nova Science Publ, 2009, pp. 113–146
- 150. Vlasova MA, Smirin BV, Pokidyshev DA, Mashina SY, Vanin AF, Malyshev IY, Manukhina EB. Mechanism of adaptation of the vascular system to chronic changes in nitric oxide level in the organism. *Bull Exp Biol Med* 2006;**142**:670–4
- 151. Gaspar A, Milhazes N, Santana L, Uriarte E, Borges F, Matos MJ. Oxidative stress and neurodegenerative diseases: looking for a therapeutic solution inspired on benzopyran chemistry. *Curr Top Med Chem* 2015;15:432–45
- 152. Cervellati C, Wood PL, Romani A, Valacchi G, Squerzanti M, Sanz JM, Ortolani B, Zuliani G. Oxidative challenge in Alzheimer's disease: state of knowledge and future needs. *J Investig Med* 2016;**64**:21–32
- 153. Lopategui Cabezas I, Herrera Batista A, Pentón Rol G. The role of glial cells in Alzheimer disease: potential therapeutic implications. *Neurologia* 2014;**29**:305–9
- 154. Benzi G, Moretti A. Age- and peroxidative stress-related modifications of the cerebral enzymatic activities linked to mitochondria and the glutathione system. *Free Radic Biol Med* 1995;**19**:77–101
- 155. Zhang YD, Zhao JJ. TFEB participates in the Aβ-induced pathogenesis of Alzheimer's disease by regulating the autophagy-lysosome pathway. DNA Cell Biol 2015;**34**:661–8
- 156. Fernández-Moriano C, González-Burgos E, Gómez-Serranillos MP. Mitochondria-targeted protective compounds in Parkinson's and Alzheimer's diseases. Oxid Med Cell Longev 2015;2015:408927

157. Ganie SA, Dar T, Bhat A, Dar K, Anees S, Masood A, Zargar MA. Melatonin: a potential antioxidant therapeutic agent for mitochondrial dysfunctions and related disorders. *Rejuvenation Res* 2016;**19**:21–40

- 158. Calabrese V, Sultana R, Scapagnini G, Guagliano E, Sapienza M, Bella R, Kanski J, Pennisi G, Mancuso C, Stella AM, Butterfield DA. Nitrosative stress, cellular stress response, and thiol homeostasis in patients with Alzheimer's disease. *Antioxid Redox Signal* 2006;8:1975–86
- 159. Maté I, Cruces J, Giménez-Llort L, De la Fuente M. Function and redox state of peritoneal leukocytes as preclinical and prodromic markers in a longitudinal study of triple-transgenic mice for Alzheimer's disease. J Alzheimers Dis 2015;43:213–26
- 160. Hritcu L, Stefan M, Brandsch R, Mihasan M. Enhanced behavioral response by decreasing brain oxidative stress to 6-hydroxy-l-nicotine in Alzheimer's disease rat model. *Neurosci Lett* 2015;**591**:41–7
- 161. Tajes M, Guivernau B, Ramos-Fernández E, Bosch-Morató M, Palomer E, Guix FX, Muñoz FJ. The pathophysiology of triose phosphate isomerase dysfunction in Alzheimer's disease. *Histol Histopathol* 2013;28:43–51
- 162. Asha Devi S, Subramanyam MV, Vani R, Jeevaratnam K. Adaptations of the antioxidant system in erythrocytes of trained adult rats: impact of intermittent hypobaric-hypoxia at two altitudes. *Comp Biochem Physiol C Toxicol Pharmacol* 2005;**140**:59–67
- 163. Rudyk OV, Vavilova HL, Strutyns'ka NA, Kotsiuruba AV, Sahach VF. Sensitivity of phenylarsineoxide-induced mitochondrial permeability transition pore in the heart of old rats during intermittent hypoxic training. *Fiziol Zh* 2004;**50**:29–37 [Ukrainian]
- 164. Gonchar O, Mankovska I. Moderate hypoxia/hyperoxia attenuates acute hypoxia-induced oxidative damage and improves antioxidant defense in lung mitochondria. *Acta Physiol Hung* 2012;99:436–46
- 165. Gonchar OA, Nosar VI, Bratus LV, Tymchenko IN, Steshenko NN, Mankovska IN. Energetic and antioxidant status of rat liver mitochondria during hypoxia-reoxygenation of different duration. *Fiziol Zh* 2015;61:35–5 [Ukrainian]
- 166. Manukhina EB, Goryacheva AV, Barskov IV, Viktorov IV, Guseva AA, Pshennikova MG, Khomenko IP, Mashina SY, Pokidyshev DA, Malyshev IY. Prevention of neurodegenerative damage to the brain in rats in experimental Alzheimer's disease by adaptation to hypoxia. *Neurosci Behav Physiol* 2010;40:737–43
- 167. Pshennikova MG, Popkova EV, Pokidyshev DA, Khomenko IP, Zelenina OM, Kruglov SV, Manukhina EB, Shimkovich MV, Goriacheva AV, Malyshev IuI. The effects of adaptation to hypoxia on the resistance to neurodegenerative disorders in the brain of rats of different genetic strains. *Vestn Ross Akad Med Nauk* 2007;2:50–5 [Russian]
- 168. Kalachev AG, El'chaniova SA, Filippova AG, Korenyak NA, Dryagina IV, Makarova IN. Use of long-term hypoxic training for secondary prevention of ischemic heart disease. *Vestnik Aritmologii* 2004;**35**:30–3 [Russian]
- 169. El'chaninova SA, Koreniak NA, Smagina IV, Pinegin LE, Varshavskii Bia. Intermittent hypoxia in the treatment of dyscirculatory encephalopathy. Zh Nevrol Psikhiatr Im S S Korsakova 2002;102:29–32 [Russian]
- 170. Mallet RT, Ryou M-G, Manukhina EB, Downey HF. Beta-adrenergic signaling and ROS: pivotal roles in intermittent, normobaric hypoxiainduced cardioprotection. In: Xi L, Serebrovskaya TV (eds). *Intermittent hypoxia: from molecular mechanisms to clinical applications*. New York: Nova Science Publ., 2009, pp. 189–212
- Vargas MR, Johnson JA. The Nrf2-ARE cytoprotective pathway in astrocytes. Expert Rev Mol Med 2009;11:e17
- 172. Niture SK, Khatri R, Jaiswal AK. Regulation of Nrf2 an update. Free Radic Biol Med 2014;66:36–44
- Dringen R, Brandmann M, Hohnholt MC, Blumrich E-M. Glutathionedependent detoxification processes in astrocytes. *Neurochem Res* 2015;40:2570–82
- 174. Estrada JA, Williams AG Jr, Sun J, Gonzalez L, Downey HF, Caffrey JL, Mallet RT. δ-Opioid receptor (DOR) signaling and reactive oxygen species (ROS) mediate intermittent hypoxia induced protection of canine myocardium. *Basic Res Cardiol* 2016;**111**:17

175. Lin L, Huang QX, Yang SS, Chu J, Wang JZ, Tian Q. Melatonin in Alzheimer's disease. Int J Mol Sci 2013;14:14575-93

- 176. Miller E, Morel A, Saso L, Saluk J. Melatonin redox activity. Its potential clinical applications in neurodegenerative disorders. *Curr Top Med Chem* 2015;15:163–9
- 177. Cardinali DP, Vigo DE, Olivar N, Vidal MF, Brusco LI. Melatonin therapy in patients with Alzheimer's disease. Antioxidants (Basel) 2014;3:245-77
- 178. Kaur C, Srinivasan KN, Singh J, Peng CM, Ling EA. Plasma melatonin, pinealocyte morphology, and surface receptors/antigen expression on macrophages/microglia in the pineal gland following a high-altitude exposure. J Neurosci Res 2002;67:533–43
- 179. Meerson FZ. Essentials of adaptive medicine: protective effects of adaptation. Moscow: Hypoxia Medical Ltd, 1994
- Ehrenbourg IV, Gorbachenkov AA. Interval hypoxic training in ischemic heart disease. *Hypox Med J* 1993;1:13–6
- Navarrete-Opazo A, Mitchell GS. Therapeutic potential of intermittent hypoxia: a matter of dose. Am J Physiol Regul Integr Comp Physiol 2014;307:R1181–97
- 182. Aleshin IA, Kots IaI, Tverdokhlib VP, Galiautdinov GS, Vdovenko LG, Zabirov MR, Meerson FZ. The nondrug treatment of hypertension patients by their adaptation to periodic hypoxia in a barochamber. *Ter Arkh* 1993;65:23–29 [Russian]
- 183. Aleshin IA, Tin'kov AN, Kots IaI, Tverdokhlib VP. Experience in treating patients with cardiovascular diseases by means of adaptation to periodic barochamber hypoxia. *Ter Arkh* 1997;69:54–8 [Russian]
- 184. Perim RR, Bonagamba LG, Machado BH. Cardiovascular and respiratory outcome of preconditioned rats submitted to chronic intermittent hypoxia. *Exp Physiol* 2015;100:1008-17
- 185. Neckár J, Sźárszoi O, Herget J, Ostádal B, Kolár F. Cardioprotective effect of chronic hypoxia is blunted by concomitant hypercapnia. *Physiol Res* 2003;52:171–5
- 186. Tin'kov AN, Aksenov VA. Effects of intermittent hypobaric hypoxia on blood lipid concentrations in male coronary heart disease patients. *High Alt Med Biol* 2002;**3**:277–82
- 187. Tomásová H, Lisý V, Trojan S, Stastný F. Effect of short-term or intermittent hypobaric hypoxia on plasma lipids in young rats. *Physiol Bohemoslov* 1987;36:361–4
- 188. Kong Z, Zang Y, Hu Y. Normobaric hypoxia training causes more weight loss than normoxia training after a 4-week residential camp for obese young adults. *Sleep Breath* 2014;18:591–7
- Kshnyaseva SI, Konstantinova OD, Tinkov AN. Hypobaric therapy in correction of cardiovascular risk factors in early postmenopausal women. *Kardiovaskulyarnaya Terapiya I Profilaktika* 2012;5:5–60 [Russian]
- 190. Urdampilleta A, Gonzalez-Muniesa P, Portillo MP, Martinez JA. Usefulness of combining intermittent hypoxia and physical exercise in the treatment of obesity. *J Physiol Biochem* 2012;68:289–304
- Lyamina NP, Pilyavsky BG. Interval hypoxic training for the treatment of cardiac rhythm disorders in patients with neurocirculatory dystonia. *Hypox Med J* 1995;1:18–20
- 192. Manukhina EB, Downey HF, Lyamina SV, Lyamina NP. Beneficial effects of adaptation to hypoxia in patients with ischemic heart disease and extrasystolic arrhythmias. J Mol Cell Cardiol 2007;42:S9
- 193. Zhou JJ, Ma HJ, Liu Y, Guan Y, Maslov LN, Li DP, Zhang Y. The antiarrhythmic effect of chronic intermittent hypobaric hypoxia in rats with metabolic syndrome induced with fructose. *Can J Physiol Pharmacol* 2015;**93**:227–32
- 194. Gao L, Chen L, Lu ZZ, Gao H, Wu L, Chen YX, Zhang CM, Jiang YK, Jing Q, Zhang YY, Yang HT. Activation of α1B-adrenoceptors contributes to intermittent hypobaric hypoxia-improved postischemic myocardial performance via inhibiting MMP-2 activation. *Am J Physiol Heart Circ Physiol* 2014;**306**:H1569–81
- 195. Zhang Y, Zhou ZN. Beneficial effects of intermittent hypobaric hypoxia on the body. *Zhongguo Ying Yong Sheng Li Xue Za Zhi* 2012;28:504–9 [Chinese]

- 196. Serebrovska TV, Shatilo VB. Remote ischemic preconditioning versus intermittent hypoxia training: a comparative analysis for cardioprotection. *Fiziol Zh* 2015;61:99–117
- 197. Pshennikova MG, Smirin BV, Bondarenko ON, Malyshev IIu, Manukhina EB. Nitric oxide storage in rats of various genetic strains and its role in the antistressor effect of adaptation to hypoxia. *Ross Fiziol Zh Im I M Sechenova* 2000;**86**:174–181 [Russian]
- 198. Zhu XH, Yan HC, Zhang J, Qiu XS, Chen L, Li SJ, Cao X, Bean JC, Chen LH, Qin XH, Liu JH, Bai XC, Mei L, Gao TM. Intermittent hypoxia promotes hippocampal neurogenesis and produces antidepressant-like effects in adult rats. *J Neurosci* 2010;**30**:12653–63
- 199. Jung ME, Simpkins JW, Wilson AM, Downey HF, Mallet RT. Intermittent hypoxia conditioning prevents behavioral deficit and brain oxidative stress in ethanol-withdrawn rats. J Appl Physiol 2008;105:510-7
- 200. Kolesnikova EE, Serebrovskaya TV. Parkinson's disease and intermittent hypoxia training. In: Xi L, Serebrovskaya TV (eds). *Intermittent hypoxia: from molecular mechanisms to clinical applications*. New York: Nova Science Publ., 2009, pp. 549–60
- 201. Gonzalez-Rothi EJ, Lee KZ, Dale EA, Reier PJ, Mitchell GS, Fuller DD. Intermittent hypoxia and neurorehabilitation. J Appl Physiol 2015;119:1455–65
- Stowe AM, Altay T, Freie AB, Gidday JM. Repetitive hypoxia extends endogenous neurovascular protection for stroke. *Ann Neurol* 2011;69:975–85
- Lin AM, Dung SW, Chen CF, Chen WH, Ho LT. Hypoxic preconditioning prevents cortical infarction by transient focal ischemia-reperfusion. Ann N Y Acad Sci 2003;993:168–78
- 204. Bigdeli MR, Rahnema M, Khoshbaten A. Preconditioning with sublethal ischemia or intermittent normobaric hyperoxia up-regulates glutamate transporters and tumor necrosis factor-alpha converting enzyme in the rat brain. J Stroke Cerebrovasc Dis 2009;**18**:336–342
- 205. Rybnikova E, Vataeva L, Tyulkova E, Gluschenko T, Otellin V, Pelto-Huikko M, Samoilov MO. Mild hypoxia preconditioning prevents impairment of passive avoidance learning and suppression of brain NGFI-A expression induced by severe hypoxia. *Behav Brain Res* 2005;**160**:107–14
- 206. Paltsyn A, Komissarova S, Dubrovin I, Kubatiev A. Increased cell fusion in cerebral cortex may contribute to poststroke regeneration. *Stroke Res Treat* 2013;**2013**:869327
- 207. Paltsyn AA, Manukhina EB, Goryacheva AV, Downey HF, Dubrovin IP, Komissarova SV, Kubatiev AA. Intermittent hypoxia stimulates formation of binuclear neurons in brain cortex-a role of cell fusion in neuroprotection? *Exp Biol Med* 2014;239:595–600
- 208. Moncayo J, de Freitas GR, Bogousslavsky J, Altieri M, van Melle G. Do transient ischemic attacks have a neuroprotective effect? *Neurology* 2000;**54**:2089–94
- 209. Meerson FZ, Pinelis VG, Koshelev VB, Riasina TV, Storozhevykh TP, Mareeva TB, Arsen'eva EN, Vorontsova EIa, Fadiukova OE. Adaptation to periodic hypoxia restricts subdural hemorrhages in audiogenic epileptiform convulsions in rats. *Biull Eksp Biol Med* 1993;**116**:572–4 [Russian]
- 210. Zhen J, Wang W, Zhou J, Qu Z, Fang H, Zhao R, Lu Y, Wang H, Zang H. Chronic intermittent hypoxic preconditioning suppresses pilocarpineinduced seizures and associated hippocampal neurodegeneration. *Brain Res* 2014;**1563**:122–30
- 211. Gao C, Wang C, Liu B, Wu H, Yang Q, Jin J, Li H, Dong S, Gao G, Zhang H. Intermittent hypoxia preconditioning-induced epileptic tolerance by upregulation of monocarboxylate transporter 4 expression in rat hippocampal astrocytes. *Neurochem Res* 2014;**39**:2160–9
- 212. Tarczyluk MA, Nagel DA, O'Neil JD, Parri HR, Tse EH, Coleman MD, Hill EJ. Functional astrocyte-neuron lactate shuttle in a human stem cell-derived neuronal network. J Cereb Blood Flow Metab 2013;33:1386–93